

## Enantioselective Synthesis and Complement Inhibitory Assay of A/B-Ring Partial Analogues of Oleanolic Acid

Haregewein Assefa, a Alison Nimrod, Larry Walkerb, c and Robert Sindelara, \*\*

<sup>a</sup>Department of Medicinal Chemistry, School of Pharmacy, University of Mississippi, University, MS 38677, USA
<sup>b</sup>National Center for Natural Product Research, School of Pharmacy, University of Mississippi, University, MS 38677, USA
<sup>c</sup>Department of Pharmacology, School of Pharmacy, University of Mississippi, University, MS 38677, USA

Received 10 November 2000; accepted 16 March 2001

Abstract—A series of oleanolic acid A/B-ring partial analogues was synthesized and tested for their complement inhibitory activity as well as cytotoxic properties. All target compounds and one intermediate exhibited moderate complement inhibitory potency. These compounds also showed cytotoxicity on malignant melanoma cell line, SK-MEL. © 2001 Elsevier Science Ltd. All rights reserved.

Complement is an essential component of the innate immune system that provides a first line defense and immune complex clearance in the blood stream.<sup>1,2</sup> Furthermore, complement plays a role in various functions of the adaptive immune response.<sup>3,4</sup> Yet overactivation of complement is implicated in various inflammatory diseases and xenotransplant rejection. 5–8 Complement inhibitors have been found to ameliorate these deleterious conditions, and a continued search for complement inhibitors has resulted in the identification of numerous natural and synthetic compounds.9-11 However, no complement inhibitor has been approved for clinical use in the US due to the lack of potency and selectivity. The triterpene natural product, oleanolic acid (3β-hydroxyolean-12-en-28-oic acid, 1) inhibits the C3-convertase of the classical pathway in vitro. 12 Oleanolic acid also inhibits complement-mediated inflammation in animal models.<sup>13</sup> Earlier, we reported the synthesis and complement inhibitory activity of various semisynthetic analogues of oleanolic acid.<sup>14</sup> To further explore the regions of the molecule that confer complement inhibitory activity, we have designed and synthesized A/Bring partial analogues of oleanolic acid. This dissection of the molecule also provides an easy access to a wider variety of analogues. The A/B-ring of oleanolic acid with its stereochemistry has been retained in the partial analogues while the carboxylic moiety has been tethered to the A/B-ring using a styrene (2a–c) or a benzyl spacer group (3a–c) (Fig. 1).

The optically pure compound 7 was synthesized using the method developed by Hagiwara and Uda. Accordingly, condensation of 2-methyl-1,3-cyclohexanedione (4) with ethyl vinyl ketone in the presence of

Figure 1. Oleanolic acid (1) and its partial analogues (2a-c and 3a-c).

0960-894X/01/\$ - see front matter  $\odot$  2001 Elsevier Science Ltd. All rights reserved. P11: S0960-894X(01)00210-4

<sup>\*</sup>Corresponding author. Tel.: +1-662-915-7101; fax: +1-662-915-5638; e-mail: sindelar@olemiss.edu

9 
$$\frac{a}{71\%}$$
  $\frac{b}{79\%}$   $\frac{cooch_3}{Ho}$   $\frac{cooch_3}{H$ 

 $\textbf{Scheme 2.} \ \ Reagents \ \ and \ \ conditions: (a) \ \ NaH/DMSO, \ \ (\textit{p}\text{-}carbomethoxybenzyl) triphenylphosphonium \ \ bromide; (b) \ \ DDQ/CH_2Cl_2/H_2O; (c) \ \ aq \ \ NaOH/CH_3OH/THF.$ 

triethylamine under reflux followed by enantioselective cyclization of the resulting triketone using L-phenylalanine (L-Phe) as a chiral auxiliary afforded the S-enantiomer 5 as a major product. The pure S-enantiomer 5 obtained by crystallization was used as a starting material for the subsequent steps. Selective protection of the unconjugated keto group of 5 using 2-ethyl-2-methyl-1,3-dioxolane in the presence of d-camphorsulfonic acid (d-CSA) and treatment of the ketal with Li/liq NH<sub>3</sub> followed by iodomethane yielded the trans-decalin 6. Reduction of the keto group of 6 using NaBH<sub>4</sub> gave the β-epimer 7 as the major product, which was separated from its α-epimer by flash chromatography to yield up to 81% of stereochemically pure 7. Protection of the hydroxyl group of compound 7 using p-methoxybenzyl chloride and NaH and deprotection of the keto group using 1 N HCl and acetic acid in tetrahydrofuran resulted in the common intermediate 8.16 Wittig reaction of 8 with (methoxymethyl)triphenylphosphonium chloride using dimsylsodium as a base followed by hydrolysis of the resulting enol ether using hydrochloric acid in tetrahydrofuran (THF) afforded the β-aldehyde 9 as the major product. 17 Even though the  $\alpha$ -epimer was formed as a minor product, column chromatographic separation gave a high yield of the pure epimer 9.

The stereochemistry of the aldehyde 9 was confirmed by 2-D NMR spectroscopic analysis. NOESY correlation

between the angular methyl and the aldehyde protons clearly indicates the  $\beta$ -orientation of the formyl group. This stereochemical assignment was supported by molecular modeling studies. <sup>18</sup> The distance between the angular methyl and the formyl group hydrogens in the  $\beta$ -epimer 9 was calculated to be as close as 2.04 Å, while the minimum distance between these two hydrogens in the  $\alpha$ -epimer was calculated to be 4.5 Å.

The synthesis of the target compounds **2a**–**c** began with the Wittig reaction of **9** with the respective triphenylphosphonium salts<sup>19</sup> using dimsylsodium as a base.<sup>20</sup> Thus, reaction of the aldehyde **9** with (*p*-carbomethoxybenzyl)triphenylphosphonium bromide gave the *trans* compound **10** as the major product, which was easily separable from its *cis*-isomer (minor product) by flash chromatography. Deprotection of the benzyl protecting group of **10** using 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (DDQ)<sup>21</sup> gave compound **11**, which on hydrolysis using NaOH yielded the compound **2a** as the final product.

Reaction of **9** with (*m*-cyanobenzyl)triphenylphosphonium bromide yielded **12** as a *cis–trans* inseparable mixture (1:3 ratio, <sup>1</sup>H NMR). Therefore, the mixture was subjected to deprotection of the hydroxyl group with DDQ to give **13**. Hydrolysis of **13** using KOH in THF and methanol under reflux yielded the hydroxy acid (85%) along with the corresponding primary amide

Scheme 3. Reagents and conditions: (a) NaH/DMSO, (*m*-cyanobenzyl)triphenylphosphonium bromide; (b) DDQ/CH<sub>2</sub>Cl<sub>2</sub>/H<sub>2</sub>O; (c) aq KOH/CH<sub>3</sub>OH/THF, Δ; (d) NaH/DMSO, (*o*-cyanobenzyl)triphenylphosphonium bromide; (e) aq KOH/ethylene glycol, Δ.

Scheme 4. Reagents and conditions: (a) LDA, methyl p-(bromomethyl)benzoate/THF; (b) NaBH<sub>4</sub>/C<sub>2</sub>H<sub>5</sub>OH; (c) NaH, CS<sub>2</sub>, CH<sub>3</sub>I/THF; (d) n-Bu<sub>3</sub>SnH/toluene,  $\Delta$ ; (e) aq NaOH/CH<sub>3</sub>OH/THF; (f) H<sub>2</sub>/Pd-C.

Scheme 5. Reagents and conditions: (a) LDA, m-cyanobenzyl bromide or o-cyanobenzyl bromide/THF; (b) NaBH<sub>4</sub>/C<sub>2</sub>H<sub>5</sub>OH; (c) NaH, CS<sub>2</sub>, CH<sub>3</sub>I/THF; (d) n-Bu<sub>3</sub>SnH/toluene,  $\Delta$ ; (e) aq KOH/ethylene glycol,  $\Delta$ ; (f) H<sub>2</sub>/Pd–C.

(15%). The pure *trans* product **2b** was obtained by crystallization from methanol. Wittig reaction of **9** with (*o*-cyanobenzyl)triphenylphosphonium bromide gave the *trans*-product **14** (>95%, <sup>1</sup>H NMR) as the major

product. Cleavage of the *p*-methoxybenzyl group of **14** using DDQ afforded the alcohol **15**. Recrystallization afforded the pure *trans*-isomer, which was subjected to hydrolysis to give the final compound **2c**.

The synthesis of 3a-c was initiated by alkylation of 8 using LDA and benzyl bromides in THF.<sup>22</sup> Therefore, alkylation of 8 with methyl p-(bromomethyl)benzoate afforded the  $\alpha$ -epimer 16 as the major product. Recrystallization from hexane afforded the pure  $\alpha$ -epimer 16. Compound 16 was also obtained by stirring with K<sub>2</sub>CO<sub>3</sub> in methanol probably due to the epimerization of the β-epimer (axial benzyl substituent) to the more stable  $\alpha$ -epimer (equatorial benzyl substituent). The stereochemistry of 16 was assigned by 2-D NMR analysis in which the NOESY correlation between the angular methyl and the proton alpha to the carbonyl group was considered as a clear indication of the α-orientation of the benzyl substituent. The keto group was successfully converted to the corresponding methylene group by reducing the ketone to the corresponding alcohol using NaBH<sub>4</sub> and removing the resultant hydroxyl group using Barton deoxygenation protocol.<sup>23</sup> Hydrolysis of the ester group of 17 by stirring with aqueous NaOH in methanol and THF afforded 18. Then the final product 3a was obtained by hydrogenolysis of the p-methoxybenzyl group of **18** using Pd–C as a catalyst.

Alkylation of 8 with m-cyanobenzyl bromide or o-cyanobenzyl bromide afforded the  $\alpha$ -epimer of compounds 19b and 19c as major products. The pure epimer ( $\alpha$ epimer) of each compound was obtained by stirring the mixture with K<sub>2</sub>CO<sub>3</sub> in methanol. The relative stereochemistry was determined using the same method employed for the stereochemical assignment of compound 16. Molecular modeling studies using the same program and methods described for 9 indicate that the distance between the angular methyl hydrogen and the hydrogen alpha to the carbonyl group is as close as 2.01 A. The carbonyl group of 19b and 19c was reduced to the corresponding methylene group using the protocol described for the conversion of compound 16 to 17. The target compounds 3b and 3c were obtained by hydrolysis of the cyano group of **20b** and **20c** using aq KOH in ethylene glycol under reflux, followed by hydrogenolysis of the *p*-methoxybenzyl group.

The compounds were bioassayed for the classical pathway complement inhibitory activity in vitro following the protocol described earlier. The cytotoxic property of the compounds was assessed in a human malignant melanoma cell line, SK-MEL. Solution of the compounds were incubated with 25,000 SK-MEL cells/well for 72 h. The number of remaining viable cells was assessed using the supravital dye, neutral red. Briefly, cells were washed with saline, incubated for 1.5 h with a 0.17% solution of neutral red in serum-free RPMI, and washed again to remove extracellular dye. Following solubilization with 0.04 N HCl in isopropanol, absorbance was read at 490 nM.

All the target compounds 2a-c and 3a-c, and the intermediate compound 21b have shown moderate complement inhibitory potency (Table 1). The lack of complement inhibitory activity with compounds 11 and 15 shows the importance of the free carboxylic group for complement inhibition. The lack of complement inhibitory activity with compounds 18 and 21c while

**Table 1.** Classical pathway complement inhibition and cytotoxicity assays of oleanolic acid and its partial analogues

| Compd | Complement inhibition $IC_{50}$ ( $\mu M^a$ ) | Cytotoxicity IC <sub>50</sub> (µM) | T.I. <sup>b</sup> |
|-------|---|------------------------------------|-------------------|
| 1     | $72.3~(\pm 5.8)$                              | 112                                | 1.55              |
| 11    | na  | na                                 | _                 |
| 2a    | $623 (\pm 34)$                                | 380                                | 0.6               |
| 15    | na  | 155                                | _                 |
| 2b    | $550 (\pm 86)$                                | 269                                | 0.5               |
| 2c    | $633(\pm 83)$                                 | 380                                | 0.6               |
| 18    | na  | 200                                |                   |
| 3a    | $610 \ (\pm 9)$                               | 378                                | 0.6               |
| 21b   | $488(\pm 11)$                                 | 180                                | 0.4               |
| 21c   | na  | 166                                |                   |
| 3b    | $601 (\pm 51)$                                | 333                                | 0.6               |
| 3c    | $616(\pm 66)$                                 | 484                                | 0.8               |

<sup>a</sup>Values are means of three experiments, standard deviation is given in parentheses (na = not active).

 $^{\hat{b}}$ In vitro therapeutic index (IC<sub>50</sub> cytotoxicity/IC<sub>50</sub> complement inhibition).

compound **21b** retains the activity may indicate that the *meta* position of the carboxylic group is more favorable for complement inhibitory activity.

All the target compounds 2a–c and 3a–c as well as the intermediates 15, 18, 21b, and 21c showed cytotoxic activity. Cytotoxicity and complement inhibitory activity appear to be dissociated as indicated by compounds 15, 18, and 21c. Although these partial analogues showed moderate complement inhibitory potency with low value of T.I., the flexible synthetic route developed should allow easy access to various analogues for further structure–activity relationship studies. It may also be possible to enhance the cytotoxicity to useful levels should the mechanism of cytotoxicity prove novel.

## Acknowledgements

This work was supported in part by the United States Department of Agriculture, Agriculture Research Service Specific Cooperative Agreement No. 58-6408-7-012.

## References and Notes

- 1. Complement, 2nd ed.; Law, S. K. A., Reid, K. B. M., Eds.; IRL: New York, 1995.
- 2. Dodds, A. Biochemist 1998, 20, 18.
- 3. Taylor, P.; Botto, M.; Walport, M. Curr. Biol. 1998, 8, R259
- 4. Carroll, M. C. Semin. Immunol. 1998, 10, 279.
- 5. Robbins, R. A.; Russ, W. D.; Rasmussen, J. K.; Clayton, M. M. Am. Rev. Respir. Dis. 1987, 135, 651.
- Kemp, P. A.; Spragg, J. H.; Brown, J. C.; Morgan, B. P.; Gunn, C. A.; Taylor, P. W. J. Clin. Lab. Immunol. 1992, 37, 147.
- 7. Homeister, J. W.; Lucchest, B. R. Annu. Rev. Pharmacol. Toxicol. 1994, 34, 17.
- 8. Rogers, J.; Griffin, W. S. T. In *Neuroinflammation: Mechanisms and Management*; Wood, P. L., Ed.; Humana: Totowa, 1998; p 177.
- 9. Hagmann, W. K.; Sindelar, R. D. Annu. Rep. Med. Chem. 1992, 27, 199.

- 10. Makrides, S. C. Pharmacol. Rev. 1998, 50, 59.
- 11. Knaus, U.; Wagner, H. Phytomedicine 1996, 3, 77.
- 12. Kapil, A.; Sharma, S. J. Pharm. Pharmacol. 1994, 46, 922.
- 13. Kapil, A.; Sharma, S. J. Pharm. Pharmacol. 1995, 47, 585.
- 14. Assefa, H.; Nimrod, A.; Walker, L.; Sindelar, R. Bioorg. Med. Chem. Lett. 1999, 9, 1889.
- 15. Hagiwara, H.; Uda, H. J. Org. Chem. 1988, 53, 2308.
- 16. Snitman, D. L.; Tsai, M.-Y.; Watt, D. S. *Synth. Commun.* **1978**, *8*, 195.
- 17. Thompson, T. N.; Sierra, M. G.; Mcchesney, J. D. J. Org. Chem. 1985, 50, 4447.
- 18. SYBYL® 6.4 Molecular Modeling Software Manual. Tripos Inc., 1699 Hanley Road, St. Louis, MO 63144-2913, USA.

- 19. The triphosphonium salt was synthesized by refluxing the respective substituted benzyl bromide with triphenylphosphine in benzene.
- 20. Greenwald, R.; Chaykovsky, M.; Corey, E. J. J. Org. Chem. 1963, 28, 1128.
- 21. Horita, K.; Yoshioka, T.; Tanaka, T.; Oikawa, Y.; Yonemitsu, O. *Tetrahedron* **1986**, *42*, 3021.
- 22. Grieco, P. A.; Ferrino, S.; Oguri, T. J. Org. Chem. 1979, 44, 2593.
- 23. Tice, C. M.; Heathcock, C. H. J. Org. Chem. 1981, 46, 9.
- 24. Srivastava, R. P.; Zhu, X.; Walker, L. A.; Sindelar, R. D. *Bioorg. Med. Chem. Lett.* **1995**, *5*, 2429.
- 25. Borenfreund, E.; Babich, H.; Martin-Alguacil, N. Toxic. In Vitro 1988, 2, 1.